

Information Sheet on the Low Glutamate Diet Pediatric Epilepsy Study

Study Overview

Glutamate has been implicated in epilepsy and seizure occurrence in both animals and humans.¹⁻³ A major hypothesis for the mechanism of action for seizure occurrence is increased levels of glutamate that result in neuronal excitability and excitotoxicity.^{4,5} Excitotoxicity in turn causes oxidative stress and increased neuroinflammation.^{6,7} Animal models have demonstrated that elevated glutamate levels can result in seizures.⁸⁻¹¹ Furthermore, monosodium glutamate (MSG), has been used for many years to induce seizures in animal models.⁸⁻¹⁶ The association between increased glutamate levels and seizures has also been supported in human studies. Studies using microdialysis have provided evidence of increased glutamate levels during interictal, as well as ictal periods, in people with medically intractable epilepsy in multiple areas of the brain, including the hippocampus and cortex.¹⁷⁻²⁰ Furthermore, research highlighting the association between seizures and an impaired blood brain barrier (BBB) suggest that this impairment leads to increased glutamate levels in the brain and as a result, excess neuronal firing.^{21,22} Since BBB permeability is common in those with epilepsy, dietary glutamate should theoretically be able to affect brain glutamate levels.

The objective of this research is to test a low glutamate diet as a potential adjunct treatment for pediatric epilepsy. The low glutamate diet reduces free glutamate consumption by removing sources of free glutamate (found in food additives and foods with naturally occurring glutamate like soy sauce and aged cheeses), while also maximizing intake of neuroprotective micronutrients, which protect against glutamate induced excitotoxicity, and antioxidants which counter the oxidative stress caused by excitotoxicity.

This is a pilot study that examines the efficacy of the low glutamate diet as a treatment for childhood epilepsy. The primary objective is to assess treatment efficacy based on seizure reduction, while secondary aims are to assess changes in seizure severity, seizure duration, and quality of life.

Background Literature

The low glutamate diet was created in the Nutritional Neuroscience Lab of Dr. Kathleen Holton. As outlined above, glutamate has been strongly implicated in the etiology of seizures, thus, reducing glutamate levels through dietary change could be of profound utility in epilepsy. While healthy individuals are normally protected against high dietary intake of glutamate, any permeability of the blood-brain barrier (BBB) can allow these higher concentrations to access the brain more easily. The BBB has been shown to become permeable from high stress, head injury, hypoxia, and infections.⁴²⁻⁵⁶ Additionally, seizures and epilepsy have been associated with impairment of the BBB.³⁸ Furthermore, a recent study found that seizure induced BBB disruption in rats resulted in an increased permeability for low and high molecular weight substances.³⁹ Increased permeability of the BBB would allow dietary glutamate to cross the BBB in much higher amounts, allowing it to potentially affect epileptic activity.

Previous Research on the Low Glutamate Diet

The low glutamate diet has previously been examined in patients with fibromyalgia and irritable bowel syndrome. Fibromyalgia patients have been shown to have higher brain glutamate levels than controls (as have migraine patients).⁴⁰⁻⁴² Our lab has shown a significant reduction in neurological symptoms among FM patients after 1 month on the low glutamate diet, with a significant return of symptoms upon challenge with MSG relative to placebo in a double-blind

placebo-controlled crossover challenge. In this study, 84% of patients had >30% of their symptoms (avg = 11 symptoms) remit after 1 month on the diet.⁴³ Symptoms which remitted included diarrhea, abdominal pain, severe fatigue, muscle pain, cognitive dysfunction, problems sleeping, paresthesia, and headache (including migraine), and eight subjects had complete symptom remission.⁴³ Furthermore, when challenged with monosodium glutamate (MSG), there was a significant return of symptoms as compared to placebo.⁴³ An improvement in chronic pain symptoms from implementation of the low glutamate diet has also been suggested in global health research on widespread chronic pain in Kenya.⁴⁴ The diet was also recently tested in veterans suffering from Gulf War Illness (GWI), which is a chronic pain condition that is also characterized by other neurological symptoms including headache/migraine, widespread chronic pain, fatigue, cognitive dysfunction, anxiety, paresthesia, and sleep issues. Data from this GWI research demonstrated similar profound improvements in neurological symptoms including overall symptom load, chronic pain (including migraine), fatigue, cognitive function (under review at *Brain and Cognition*) and mood disorders (being submitted to *Biological Psychiatry*).⁴⁵ There is strong biological plausibility for why there is improvement in these symptoms, as all are known to be associated with abnormal glutamatergic neurotransmission. Of important note, the GWI research has had to restrict access to those who also suffer from seizures due to the challenge portion of the study, where subjects are challenged with MSG versus placebo, which increases risk to subjects with seizures. Since the low glutamate diet has been found to help in other neurological conditions, and glutamate has been implicated in epilepsy and seizure activity, it is highly plausible that the diet could also be effective in epilepsy patients. This study will be the first time the low glutamate diet has been tested in epilepsy. It is important to note that for this epilepsy research, we are removing the challenge with MSG versus placebo we have used previously, to remove any risk of seizure induction.

Research Plan and Methods

The proposed study is a randomized controlled clinical trial to supply evidence of the feasibility, tolerability, and efficacy of a low glutamate diet as a treatment for refractory epilepsy. A wait-listed control group is being used for comparison to subjects' "diet-as-usual," while ethically allowing every person to also have access to the intervention at some point in time.

We are recruiting 40 children with refractory epilepsy. To be included in the study, children must be between the ages of 2 and 21, be experiencing 1 or more seizures per week, have had unsuccessful seizure control with a minimum of 2 anticonvulsant drugs by the time of enrollment, and be willing to keep all medications and Vagus nerve stimulator or Responsive neurostimulation device settings constant during the study period. Additionally, all seizure medications and Vagus nerve stimulator or Responsive neurostimulation device settings will have to have remained constant for 30 days and any previously attempted dietary therapies stopped prior to joining the study. Exclusion criteria includes formula-fed patients, those with inborn errors of metabolism, and non-English speaking households.

Families will be asked to record a seizure diary for a one-month baseline period to establish baseline seizure frequency, duration, and severity. During the last week of the month, families will be asked to complete baseline measures including a seizure assessment form, the Quality of Life in Childhood Epilepsy Questionnaire, the Autism Quotient, the National Institute for Children's Health Quality Vanderbilt Assessment Scale, a Symptom Checklist, a 3-day food diary, Food Frequency Questionnaire (FFQ), and cognitive testing. Once baseline measures are

complete, families will then be randomized to the active intervention group or a one-month waitlisted control group.

Participants randomized to start the diet immediately will receive intensive online nutrition training with Dr. Holton and will be emailed diet materials to support them in following the low-glutamate diet for 1 month. The diet materials include information on food additives to avoid, naturally occurring sources of free glutamate, tricky places these additives are found, shopping lists, recipes (including ways to make healthy food more palatable to children), lists of foods highest in each micronutrient, and a high antioxidant food list. They will have access to Dr. Holton throughout the month to answer questions as they arise. Study measures will again be completed at the end of the one-month period for both active intervention and wait-listed control groups. If waitlisted, controls will then have the opportunity to undergo dietary training, follow the diet for the following month, and then will have all outcomes reassessed.

Medical Oversight

Dr. Kao will serve as medical oversight for the study. Dr. Amy Kao is a pediatric neurologist and epileptologist. She was previously the director of the Dietary Therapies for Epilepsy Clinic at Children's National Medical Center in Washington, D.C. and currently works for the FDA. This research is not related to the FDA.

Contact information

For more information regarding the study please visit clinicaltrials.gov or contact us. For more information on glutamate, dietary glutamate, and epilepsy, please contact Dr. Holton.

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References

1. Barker-Haliski M, White HS. Glutamatergic Mechanisms Associated with Seizures and Epilepsy. *Cold Spring Harb Perspect Med*. 2015;5(8):a022863. doi:10.1101/cshperspect.a022863
2. Epilepsy G, Chapman AG. Glutamate and Glutamine in the Brain. *J Nutr*. 2000;130:1043-1045. https://watermark.silverchair.com/4w04t0s1043.pdf?token=AQECAHi208BE49Ooan9kkhW_Ercy7Dm3ZL_9Cf3qfKAc485ysgAAAawwggGoBgkqhkiG9w0BBwagggGZMIIBIQIBADCCAY4GCSqGSib3DQEHAATAeBglghkgBZQMEAS4wEQQMQZAp8j0y-qFg5j38AgEQgIIBX266bD7AlXqHODPqntV72SWGpkm5Jde_Ob9S1KfB9px. Accessed February 2, 2018.
3. Meldrum BS. The role of glutamate in epilepsy and other CNS disorders. *Neurology*. 1994;44(11 Suppl 8):S14-23. <http://www.ncbi.nlm.nih.gov/pubmed/7970002>. Accessed November 27, 2019.
4. Haglid KG, Wang S, Qiner Y, Hamberger A. Excitotoxicity. Experimental correlates to human epilepsy. *Mol Neurobiol*. 1994;9(1-3):259-263. doi:10.1007/BF02816125
5. Coulter DA, Eid T. Astrocytic regulation of glutamate homeostasis in epilepsy. *Glia*. 2012;60(8):1215-1226. doi:10.1002/glia.22341
6. Viviani B, Boraso M, Marchetti N, Marinovich M. Perspectives on neuroinflammation and excitotoxicity: A neurotoxic conspiracy? *Neurotoxicology*. 2014;43:10-20. doi:10.1016/J.NEURO.2014.03.004
7. Nguyen D, Alavi M V, Kim K-Y, et al. A new vicious cycle involving glutamate excitotoxicity, oxidative stress and mitochondrial dynamics. *Cell Death Dis*. 2011;2(12):e240-e240. doi:10.1038/cddis.2011.117
8. Arauz-Contreras J, Feria-Velasco A. Monosodium-l-glutamate-induced convulsions—I. Differences in seizure pattern and duration of effect as a function of age in rats. *Gen Pharmacol Vasc Syst*. 1984;15(5):391-395. doi:10.1016/0306-3623(84)90036-3
9. Beas-Zárate C, Schliebs R, Morales-Villagran A, Feria-Velasco A. Monosodium L-glutamate-induced convulsions: changes in uptake and release of catecholamines in cerebral cortex and caudate nucleus of adult rats. *Epilepsy Res*. 4(1):20-27. <http://www.ncbi.nlm.nih.gov/pubmed/2753019>. Accessed January 13, 2019.
10. López-Pérez SJ, Ureña-Guerrero ME, Morales-Villagrán A. Monosodium glutamate neonatal treatment as a seizure and excitotoxic model. *Brain Res*. 2010;1317:246-256. doi:10.1016/j.brainres.2009.12.054
11. Trentini GP, Botticelli A, Botticelli CS. Effect of monosodium glutamate on the endocrine glands and on the reproductive function of the rat. *Fertil Steril*. 1974;25(6):478-483. <http://www.ncbi.nlm.nih.gov/pubmed/4835604>. Accessed January 13, 2019.
12. Ureña-Guerrero ME, Orozco-Suárez S, López-Pérez SJ, Flores-Soto ME, Beas-Zárate C. Excitotoxic neonatal damage induced by monosodium glutamate reduces several GABAergic markers in the cerebral cortex and hippocampus in adulthood. *Int J Dev Neurosci*. 2009;27(8):845-855. doi:10.1016/j.ijdevneu.2009.07.011
13. Gudiño-Cabrera G, Ureña-Guerrero ME, Rivera-Cervantes MC, Feria-Velasco AI, Beas-Zárate C. Excitotoxicity Triggered by Neonatal Monosodium Glutamate Treatment and Blood–Brain Barrier Function. *Arch Med Res*. 2014;45(8):653-659. doi:10.1016/j.arcmed.2014.11.014
14. Beas-Zárate C, Arauz-Contreras J, Velazquez A, Feria-Velasco A. Monosodium l-glutamate-induced convulsions—II. Changes in catecholamine concentrations in various brain areas of adult rats. *Gen Pharmacol Vasc Syst*. 1985;16(5):489-493. doi:10.1016/0306-3623(85)90009-6
15. Synowiec AS, Singh DS, Yenugadhati V, Valeriano JP, Schramke CJ, Kelly KM. Ketamine use in the treatment of refractory status epilepticus. *Epilepsy Res*. 2013;105(1-2):183-188. doi:10.1016/j.epilepsyres.2013.01.007
16. Peñafiel R, Cremades A, Monserrat F, Puellas L. Monosodium glutamate induced convulsions in rats: Influence of route of administration, temperature and age. *Amino Acids*. 1991;1(1):81-89. doi:10.1007/BF00808094
17. DURING D, Spencer D. Extracellular hippocampal glutamate and spontaneous seizure in the

- conscious human brain. *Lancet*. 1993;341(8861):1607-1610. doi:10.1016/0140-6736(93)90754-5
18. Cavus I, Kasoff WS, Cassaday MP, et al. Extracellular metabolites in the cortex and hippocampus of epileptic patients. *Ann Neurol*. 2005;57(2):226-235. doi:10.1002/ana.20380
 19. Çavuş I, Romanyshyn JC, Kennard JT, et al. Elevated basal glutamate and unchanged glutamine and GABA in refractory epilepsy: Microdialysis study of 79 patients at the yale epilepsy surgery program. *Ann Neurol*. 2016;80(1):35-45. doi:10.1002/ana.24673
 20. Ronne-Engström E, Hillered L, Flink R, Spännare B, Ungerstedt U, Carlson H. Intracerebral Microdialysis of Extracellular Amino Acids in the Human Epileptic Focus. *J Cereb Blood Flow Metab*. 1992;12(5):873-876. doi:10.1038/jcbfm.1992.119
 21. Marchi N, Granata T, Ghosh C, Janigro D. Blood-brain barrier dysfunction and epilepsy: pathophysiologic role and therapeutic approaches. *Epilepsia*. 2012;53(11):1877-1886. doi:10.1111/j.1528-1167.2012.03637.x
 22. van Vliet EA, Aronica E, Gorter JA. Blood–brain barrier dysfunction, seizures and epilepsy. *Semin Cell Dev Biol*. 2015;38:26-34. doi:10.1016/j.semcdb.2014.10.003
 23. Shanker Sharma H, Cervós-Navarro J, Kumar Dey P. Increased blood-brain barrier permeability following acute short-term swimming exercise in conscious normotensive young rats. *Neurosci Res*. 1991;10(3):211-221. doi:10.1016/0168-0102(91)90058-7
 24. Belova TI, JONSSON G. Blood-brain barrier permeability and immobilization stress. *Acta Physiol Scand*. 1982;116(1):21-29. doi:10.1111/j.1748-1716.1982.tb10594.x
 25. Price L, Wilson C, Grant G. Blood–Brain Barrier Pathophysiology following Traumatic Brain Injury. 2016. <https://www.ncbi.nlm.nih.gov/books/NBK326726/>. Accessed November 27, 2019.
 26. Škultétyová I, Tokarev D, bulletin DJ-B research, 1998 undefined. Stress-induced increase in blood–brain barrier permeability in control and monosodium glutamate-treated rats. *Elsevier*. <https://www.sciencedirect.com/science/article/pii/S0361923097003353>. Accessed November 27, 2019.
 27. Lochhead JJ, Ronaldson PT, Davis TP. Hypoxic Stress and Inflammatory Pain Disrupt Blood-Brain Barrier Tight Junctions: Implications for Drug Delivery to the Central Nervous System. *AAPS J*. 2017;19(4):910-920. doi:10.1208/s12248-017-0076-6
 28. Yeh W-L, Lu D-Y, Lin C-J, Liou H-C, Fu W-M. Inhibition of hypoxia-induced increase of blood-brain barrier permeability by YC-1 through the antagonism of HIF-1alpha accumulation and VEGF expression. *Mol Pharmacol*. 2007;72(2):440-449. doi:10.1124/mol.107.036418
 29. Alluri H, Wiggins-Dohlvik K, Davis ML, Huang JH, Tharakan B. Blood–brain barrier dysfunction following traumatic brain injury. *Metab Brain Dis*. 2015;30(5):1093-1104. doi:10.1007/s11011-015-9651-7
 30. Chodobski A, Zink BJ, Szmydynger-Chodobska J. Blood-brain barrier pathophysiology in traumatic brain injury. *Transl Stroke Res*. 2011;2(4):492-516. doi:10.1007/s12975-011-0125-x
 31. Sarmiento A, Borges N, Azevedo I. Adrenergic influences on the control of blood-brain barrier permeability. *Naunyn Schmiedebergs Arch Pharmacol*. 1991;343(6):633-637. doi:10.1007/bf00184295
 32. Esposito P, Chandler N, Kandere K, et al. Corticotropin-Releasing Hormone and Brain Mast Cells Regulate Blood-Brain-Barrier Permeability Induced by Acute Stress. *J Pharmacol Exp Ther*. 2002;303(3):1061-1066. doi:10.1124/jpet.102.038497
 33. Esposito P, Gheorghie D, Kandere K, et al. Acute stress increases permeability of the blood-brain-barrier through activation of brain mast cells. *Brain Res*. 2001;888(1):117-127. doi:10.1016/s0006-8993(00)03026-2
 34. Varatharaj A, Galea I. The blood-brain barrier in systemic inflammation. *Brain Behav Immun*. 2017;60:1-12. doi:10.1016/J.BBI.2016.03.010
 35. Chaudhuri JD. Blood brain barrier and infection. *Med Sci Monit*. 6(6):1213-1222. <http://www.ncbi.nlm.nih.gov/pubmed/11208482>. Accessed November 27, 2019.
 36. Kaur C, Ling E. Blood Brain Barrier in Hypoxic-Ischemic Conditions. *Curr Neurovasc Res*. 2008;5(1):71-81. doi:10.2174/156720208783565645

37. Friedman A, Kaufer D, Shemer J, Hendler I, Soreq H, Tur-Kaspa I. Pyridostigmine brain penetration under stress enhances neuronal excitability and induces early immediate transcriptional response. *Nat Med.* 1996;2(12):1382-1385. doi:10.1038/nm1296-1382
38. Oby E, Janigro D. The Blood-Brain Barrier and Epilepsy. *Epilepsia.* 2006;47(11):1761-1774. doi:10.1111/j.1528-1167.2006.00817.x
39. Vazana U, Veksler R, Pell GS, et al. Glutamate-Mediated Blood-Brain Barrier Opening: Implications for Neuroprotection and Drug Delivery. *J Neurosci.* 2016;36(29):7727-7739. doi:10.1523/JNEUROSCI.0587-16.2016
40. Harris RE, Sundgren PC, Craig AD, et al. Elevated insular glutamate in fibromyalgia is associated with experimental pain. *Arthritis Rheum.* 2009;60(10):3146-3152. doi:10.1002/art.24849
41. Pyke TL, Osmotherly PG, Baines S. Measuring Glutamate Levels in the Brains of Fibromyalgia Patients and a Potential Role for Glutamate in the Pathophysiology of Fibromyalgia Symptoms. *Clin J Pain.* 2017;33(10):944-954. doi:10.1097/AJP.0000000000000474
42. Holton K. The role of diet in the treatment of fibromyalgia. *Pain Manag.* 2016;6(4):317-320. doi:10.2217/pmt-2016-0019
43. Holton KF, Taren DL, Thomson CA, Bennett RM, Jones KD. The effect of dietary glutamate on fibromyalgia and irritable bowel symptoms. *Clin Exp Rheumatol.* 2012;30(6 Suppl 74):10-17. <http://www.ncbi.nlm.nih.gov/pubmed/22766026>. Accessed January 13, 2019.
44. Holton KF, Ndege PK, Clauw DJ. Dietary correlates of chronic widespread pain in Meru, Kenya. *Nutrition.* 2018;53:14-19. doi:10.1016/j.nut.2018.01.016
45. Holton KF, Kirkland AE, Baron M, et al. The Low Glutamate Diet Effectively Improves Pain and Other Symptoms of Gulf War Illness. *Nutrients.* 2020;12(9):2593. doi:10.3390/nu12092593